Short Communication

$GSK\textbf{-}3\beta \text{ expression in cerebrospinal fluid in} \\ Parkinson's \text{ disease} \\$

Jose Antonio Molina^{1,2,3}, Desiree Antequera^{2,3}, Alvaro Sanchez-Ferro^{1,2,3}, Felix Bermejo-Pareja^{1,2,3} and Eva Carro^{2,3,*}

¹Neurology Service, Hospital 12 de Octubre, Madrid, Spain,

²Center for Networker Biomedical Research in Neurodegenerative Diseases (CIBERNED), Spain

³Neuroscience Group, Instituto de Investigación Hospital 12 de Octubre, Madrid, 28041, Spain

ABSTRACT

We examined the expression of glycogen synthase kinase-3beta (GSK-3 β) in the cerebrospinal fluid (CSF) of Parkinson's disease (PD) patients. GSK-3 β is a critical intermediate in pro-apoptotic signaling cascades that are associated with neuro-degenerative diseases, including PD. Because CSF contains proteins and metabolites of brain origin, and GSK-3 β levels are increased in brains of PD patients, we decided to measure GSK-3 β levels in CSF samples from PD patients. We observed that neither GSK-3 β nor pGSK-3 β (Ser9) was altered in PD patients. Our results suggest that, although GSK-3 β plays a role in the pathogenesis of PD, its CSF levels were unchanged.

KEYWORDS: cerebrospinal fluid, Parkinson's disease, glycogen synthase kinase-3beta, risk factor

INTRODUCTION

Parkinson's disease (PD), a progressive neurodegenerative disease of unknown etiology [1], is the second most common neurological disorder characterized by motor and behavioral disturbances caused by the degeneration of dopaminergic neurons [2, 3]. Alpha-synuclein (α -syn), a presynaptic protein, is causal in the genesis of PD [2, 4], and gene duplication and triplication of α -Syn are found in sporadic and early onset forms of PD [5]. α -Syn becomes insoluble, self-aggregates and accumulates into intra-neuronal inclusion bodies [4, 6, 7, 8]. A recent study indicates that α -Syn induces hyperphosphorylated tau through specific activation and recruitment of glycogen synthase kinase 3 β (GSK-3 β), a kinase known to hyperphosphorylate tau at distinct sites in Alzheimer's disease, which itself becomes activated through autophosphorylation at Tyr216 [9].

GSK-3ß is a ubiquitous serine/threonine protein kinase highly abundant in brain tissue, which plays a key role in neural development and neuron survival, and an increase in GSK-3B activity precedes the induction of apoptosis [10]. Elevated levels of GSK-3 β were seen in the striatum [9, 11], and peripheral blood lymphocytes [12] from PD patients. These findings were also underscored by a report in which two functional single nucleotide polymorphisms have been reported in PD brains, resulting in increased phosphorylation of tau and interaction of GSK-3^β with tau [13]. All these observations suggest that this kinase may be involved in the pathogenic progress of the human PD. We further tested whether the increase in GSK-3^β levels is also reflected in the cerebrospinal fluid (CSF) of PD patients.

^{*}Corresponding author: carroeva@h12o.es

MATERIAL AND METHODS

Human samples

CSF samples from PD patients [n = 29 (12 females)]and 17 males), age = 62.97 ± 2.06 years (data are expressed as mean \pm s.e.m.)] and healthy control subjects [(n = 22 (12 females and 10 males)]age = 58.33 ± 3.8 years)] were obtained by lumbar puncture, collected in a glass tube, and immediately frozen and stored at -80°C. All the samples were obtained from the Neurology Service of the Hospital Universitario 12 de Octubre (Madrid, Spain). All PD cases were diagnosed under the criteria of probable PD [14]. The control group was formed of family members or caregivers of the PD patients, who all had a clinical interview with a senior neurologist that showed a completely normal cognitive and functional level. Half ml of each CSF sample was concentrated by lyophylization in a Speed-Vac to about 0.1 ml.

Immunoblot analysis

CSF protein immunoblotting was performed using previously describe methods [15]. In brief, all samples were resolved by 10% SDSpolyacrylamide gel electrophoresis and transferred to nitrocellulose membranes (Bio-Rad) by electroblotting. Primary antibodies were: mouse monoclonal anti-GSK-3 β (Santa Cruz Biotechnology, Inc), and rabbit polyclonal anti-pSer9 GSK-3 β (Cell Signaling Technology, Inc.). Secondary antibodies were: goat anti-mouse, and goat antirabbit HRP-conjugated (Biorad Laboratories). Densitometric analysis was performed using ImageJ software (NIH Image).

Statistical analysis

Data were analysed with SPSS for Windows (version 15.0). Results were expressed as mean \pm s.e.m. and statistically analyzed by ANOVA followed by a Tukey-Kramer test, and Mann-Whitney *U*-test analysis when appropriate. The differences were considered to be significant at p<0.05. The Spearman rank correlation was used for correlation analyses.

RESULTS AND DISCUSSION

Using Western-blot assays, we observed typical pattern of GSK-3 β in human CSF. Neither GSK-3 β nor pSer9 GSK-3 β (Figure 1A, and B) expression was altered in CSF from PD patients compared with control subjects.

We analyzed whether there was an association between GSK-3 β and pSer9 GSK-3 β expression and sex, age and onset. We found that both GSK-3 β and pSer9 GSK-3 β expression correlate with age (Spearman rank correlation R=-0.631, p = 0.002, and R = -0.497, p = 0.019, respectively) but only in the control group.

We here describe novel findings describing GSK-3 β status in CSF from PD patients. Despite the fact that the cause of PD remains unknown, α -syn has been classically linked to PD, as the main component of Lewy bodies [16]. Phosphorylated GSK-3 β , α -syn, tau, and other proteins are involved in the pathogenesis of several neurodegenerative diseases [14]. It has been demonstrated that p-GSK-3 β is central to the development of the tauopathic process in PD models, and that blockade of the kinase with specific inhibitors

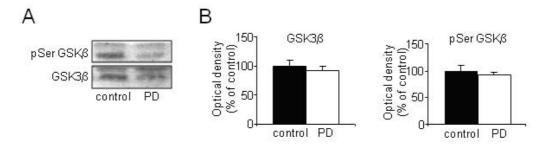


Figure 1. (A) Expression of pSer GSK-3 β and GSK-3 β are unchanged in CSF of PD patients compared with healthy controls. Representative western blots are shown. (B) Densitometric measurements confirm these data. (Data are expressed as mean \pm s.e.m., n = 22 for control group, n = 29 for PD group. PD = Parkinson's disease).

prevented tauopathic changes and cell death in mesencephalic neurons and cultured cells [17].

To our knowledge, no studies about the assessment of GSK-3 β levels in CSF from PD patients have been published. While we have not found significant differences between PD patients and controls, the role of GSK-3 β in the cellular damage in PD is well established [9, 11]. Despite the fact that GSK-3 β is up-regulated in cellular populations, including neurons [9] and blood lymphocytes [12] from PD patients, we hypothesized that its secretion to peripheral fluids, including CSF, could be reduced. However, we found that in CSF, GSK-3 β levels in PD patients were similar to those observed in healthy subjects.

ACKNOWLEDGMENTS

This work was supported by grants from Fondo de Investigación Sanitaria (FIS) (PI060155, PI0901636), Fundación Investigación Médica Mutua Madrileña (2008.93),CIBERNED, Fundación Neurociencias y Envejecimiento. This work was made possible by the generous participation of the patients, the control subjects, and their families. We thank Agnieszka Krzyzanowska, PhD, for the careful revision of this manuscript.

FINANCIAL DISCLOSURE/CONFLICT OF INTEREST

The authors report no conflicts of interest.

This work was supported by grants from Fondo de Investigación Sanitaria (FIS) (PI060155, PI0901636), Fundación Investigación Médica Mutua Madrileña (2008.93), CIBERNED, Fundacion Neurociencias y Envejecimiento.

REFERENCES

- 1. Pollanen, M. S., Dickson, D. W. and Bergeron, C. 1993, J. Neuropathol. Exp. Neurol., 52, 183-191.
- Forno, L. S. 1996, J. Neuropathol. Exp. Neurol., 55, 259-272.
- 3. de Lau, L. M. and Breteler, M. M. 2006, Lancet Neurol., 5, 525-535.
- 4. Spillantini, M. G., Crowther, R. A., Jakes, R., Hasegawa, M. and Goedert, M.

1998, Proc. Natl. Acad. Sci. USA, 95, 6469-6473.

- Singleton, A. B., Farrer, M., Johnson, J., Singleton, A., Hague, S., Kachergus, J., Hulihan, M., Peuralinna, T., Dutra, A., Nussbaum, R., Lincoln, S., Crawley, A., Hanson, M., Maraganore, D., Adler, C., Cookson, M. R., Muenter, M., Baptista, M., Miller, D., Blancato, J., Hardy, J. and Gwinn-Hardy, K. 2003, Science, 302, 841.
- Masliah, E., Rockenstein, E., Veinbergs, J., Mallory, M., Hashimoto, M., Takeda, A., Sagara, Y., Sisk, A. and Mucke, L. 2000, Science, 287, 1265-1269.
- El-Agnaf, O. M. A., Jakes, R., Curran, M. D. and Wallace, A. 1998, FEBS. Letts., 440, 67-70.
- Gosavi, N., Lee, H. J., Lee, J. S., Patel, S. and Lee, S. J. 2002, J. Biol. Chem., 277, 48984-48992.
- Duka, T., Duka, V., Joyce, J. N. and Sidhu, A. 2009, FASEB J., 23, 2820-2830.
- 10. Jope, R. S. and Johnson, G. V. 2004, Trends Biochem. Sci., 29, 95-102.
- 11. Nagao, M. and Hayashi, H. 2009, Neurosci. Lett., 449, 103-107.
- Armentero, M. T., Sinforiani, E., Ghezzi, C., Bazzini, E., Levandis, G., Ambrosi, G., Zangaglia, R., Pacchetti, C., Cereda, C., Cova, E., Basso, E., Celi, D., Martignoni, E., Nappi, G. and Blandini, F. 2011, Neurobiol. Aging, 32, 2142-2151.
- Kwok, J. B., Hallupp, M., Loy, C. T., Chan, D. K., Woo, J., Mellick, G. D., Buchanan, D. D., Silburn, P. A., Halliday, G. M. and Schofield, P. R. 2005, Ann. Neurol., 58, 829-839.
- 14. Gelb, D. J., Oliver, E. and Gilman, S. 1999, Arch. Neurol., 56, 33-39.
- Kozlovsky, N., Regenold, W. T., Levine, J., Rapoport, A., Belmaker, R. H. and Agam, G. 2004, J. Neural. Transm., 111, 1093-1098.
- 16. Petit-Paitel, A. 2010, Medicine Sciences, 26, 516-521.
- Kozikowski, A. P., Gaisina, I. N., Petukhov, P. A., Sridhar, J., King, L. T., Blond, S. Y., Duka, T., Rusnak, M. and Sidhu, A. 2006, Chem. Med. Chem., 1, 256-266.