

Original Communication

# Rare sugar **D**-psicose suppresses glycemic response after ingestion of various confections in healthy subjects

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# ABSTRACT

D-Psicose, a C-3 epimer of D-fructose, is a "rare sugar" present in small amounts in nature products. We investigated whether D-psicose suppresses the glycemic response after ingestion of various confections in healthy subjects. In Experiment 1, 22 male and 21 female subjects were randomly divided into 3 groups: marshmallow, fried cookie and chocolate groups. The test meals were these 3 confections containing 45 g of carbohydrate and hot coffee with 5 g of D-psicose or sucrose. The increase in blood glucose concentration after intake of various confections was significantly lower with simultaneous intake of coffee and D-psicose than with that of coffee with sucrose. In Experiment 2, 4 male and 5 female subjects ate the cake, almond jelly, baked cookie, fried cookie, and ganache containing 5 g of D-psicose or sucrose as raw materials. The increase in plasma glucose concentration was significantly lower after intake of almond jelly and ganache containing D-psicose than those containing sucrose. The postprandial plasma glucose concentration did not differ between intake of confections containing D-psicose and sucrose cooked at high temperature. These results suggested that D-psicose is effective for the hypoglycemic response as a functional food material. However, this effect may be suppressed by high temperature cooking. Care is necessary when D-psicose is used as a food material.

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**KEYWORDS:** D-psicose, plasma glucose, plasma insulin, confection, healthy subject

# INTRODUCTION

D-Psicose (D-ribo-2-hexulose), a C-3 epimer of D-fructose, is a "rare sugar" present in small quantities in commercial mixtures of D-glucose and D-fructose obtained from hydrolysis of sucrose or isomerisation of D-glucose [1]. D-Psicose is also present in processed cane and beet molasses [2], and is found in wheat [3], Itea plants [4], and in the antibiotic psicofranine [5]. Due to the very small amounts of D-psicose in natural products, few studies have examined D-psicose as food or nutrient. In 2004, we developed a new method for producing D-psicose enzymatically on a large scale [6], making it possible to conduct such nutritional studies. We have since demonstrated that D-psicose is a sweet monosaccharide that provides no energy to growing rats [7] and that it has little toxicity in rats [8, 9]. With respect to safety from a clinical viewpoint, the maximum non-effective level of D-psicose causing diarrhea in human subjects was estimated as 0.55 g per kg body weight [10].

Examining the effects of D-psicose on glucose and lipid metabolism, we found that D-psicose leads to less intra-abdominal fat accumulation than D-glucose and D-fructose in rats [11]. In addition, we have suggested that supplemental D-psicose can lower plasma glucose levels [12]. We have reported that D-psicose inhibits intestinal  $\alpha$ -glucosidase and suppresses the glycemic response after ingestion of disaccharides (sucrose, maltose or maltodextrin) in rats [13, 14] and

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human [15]. Recently, Toyoda *et al.* [16] suggested that D-psicose can prevent postprandial hyperglycemia by improving the translocation of glucokinase from the nucleus to the cytoplasm in the liver of diabetic rats. D-Psicose is expected to have a beneficial effect in the control of blood glucose levels in type 2 diabetes.

One of the indicated effects D-psicose is suppression of postprandial blood glucose elevation [11-15], which is helpful in reducing the risk of lifestyle-related diseases, such as type 2 diabetes. However, we have not examined the effects of D-psicose on blood glucose level after ingesting cooked meals in human subjects. In this study, we investigated whether D-psicose suppresses the glycemic response after ingestion of various confections in healthy subjects.

### MATERIALS AND METHODS

Two experiments were conducted in 52 healthy volunteers aged 20-23 recruited from Kagawa University. The subjects were determined to be free of disease by a medical examination before the study. None of the subjects were using illegal drugs or taking medications that affect blood glucose level. Volunteers were fully informed of the objective of the study, the test methods, expected adverse reactions and other related matters. Before the study started, written consent was obtained from the subjects. The study protocol and the implementation complied with the spirit of the Declaration of Helsinki in 1995 as revised in Edinburgh 2000. This trial was carried out with the approval of the Ethical Committee of Kagawa University (Approval number: H20-33).

# Experiment 1 Effects of D-psicose used as a sweetener on postprandial blood glucose level

#### Subjects and test meals

Twenty-two male and 21 female subjects were randomly divided into 3 groups: marshmallow, fried cookie and chocolate groups (Table 1). These confections were purchased from Maruyoshi Center Inc., Kagawa, Japan. The test meals were these 3 confections containing 45 g of carbohydrate and hot coffee with 5 g of D-psicose or sucrose. The compositions of test meals are shown in Table 2. The hot coffee was made from 2 g of powdered instant coffee (Nescafe, Nestle

#### **Experimental design**

Two meal load tests per subject were conducted under a randomized single blind study design. During the period of the study, each subject maintained a normal life style and ate ad libitum except for the day before the experiment, on which each subject ate the same dinner (800 kcal and 700 kcal for male and female subjects, respectively) at 19:00. After fasting for overnight, blood glucose concentration was first measured at 08:00 with a portable glucose analyzer (Glucocard G<sup>+</sup>, Arkray Inc., Kyoto, Japan). Soon after the first blood measurement, the each group of subjects took test meals within 5 min. The subjects then rested for 120 min. During the rest, the blood glucose concentrations were measured at 30, 60, 90 and 120 min after intake of test meals containing D-psicose or sucrose. The 2 meal load tests (D-psicose or sucrose) per subjects in each group were performed at intervals of at least 1 week. All procedures were performed in the experimental room under the same conditions (temperature: 22 degrees Celsius; humidity: 60%).

# Experiment 2 Effects of D-psicose used as a raw material on postprandial plasma glucose and insulin levels

#### Subjects and test meals

Four male and 5 female subjects participated in this experiment, as shown in Table 1. The compositions of test meals are shown in Table 2. The cake, almond jelly, baked cookie, fried cookie and ganache were made with or without 5g of D-psicose according to the recipe [17]. These confections were made by substitution of D-psicose in place of sucrose as a raw material.

#### **Experimental design**

The 10 meal load tests per subject were conducted under a randomized single blind study design. During the period of the study, each subject maintained a normal life style and ate *ad libitum* except for the day before the experiment, on which each subject ate the same dinner (800 kcal and 700 kcal for male and female subjects, respectively) at 19:00. After fasting for overnight,

Carrows	Experiment 1						Experi	Experiment 2	
Gloups	Marshmallow		Fried cookie		Chocolate				
	Male	Female	Male	Female	Male	Female	Male	Female	
n	7	7	8	7	7	7	4	5	
Age (y)	$20.7 \pm 0.2$	20.6±0.2	20.5±0.2	20.3±0.3	21.2±0.3	20.1±0.1	23.0±0.9	22.5±0.8	
Height (cm)	171.0±1.1	$154.0{\pm}1.0$	169.0±1.7	156.1±1.3	170.2±1.0	157.2±1.2	173.9±1.1	158.1±1.1	
Weight (kg)	62.2±1.3	50.2±1.1	60.4±1.9	50.6±1.4	64.8±1.6	51.2±1.4	73.9±2.1	54.6±2.9	
BMI (kg/m <sup>2</sup> )	21.3±1.9	21.2±1.6	21.1±2.1	20.8±1.4	22.3±1.8	20.7±1.2	24.1±1.7	20.7±1.2	

Table 1. Characteristics of subjects.

Values are means  $\pm$  SE.

	Weight	Fat	Protein	Carbohydrate	Energy
	(g)	(g)	(g)	(g)	(kcal)
Experiment 1*					
Marshmallow	56.0	0.2	1.8	45.0	189.0
Fried cookie	62.0	9.5	4.1	45.0	283.0
Chocolate	80.0	28.0	4.2	45.0	449.0
Experiment 2 <sup>#</sup>					
Cake	117.5	7.8	5.1	50.0	290.6
Almond Jelly	96.0	10.0	20.0	50.0	370.0
Baked cookie	70.9	16.1	4.8	50.0	364.1
Fried cookie	79.2	21.5	5.6	50.0	415.9
Ganache	100.0	30.0	7.9	50.0	501.6

Table 2. Composition of test meals.

\*Each confection was taken with coffee (150 ml) with sucrose (5 g) or D-psicose (5 g). \*Each confection was made with or without D-psicose (5 g) as a raw material.

blood was first collected at 08:00. Soon after the first blood collection, subjects ate the test meals within 5 min. Peripheral blood (180  $\mu$ L) was collected in heparin-coated capillaries to obtain plasma at 30, 60, 90 and 120 min after meal intake. All procedures were performed in the experimental room under the same conditions (temperature: 22 degrees Celsius; humidity: 60%).

#### Measurements

Plasma glucose and insulin concentrations were determined by be using commercial kits (Glucose CII-Test Wako, Wako Pure Chemical Industries, Ltd., Osaka, Japan; Mercodia Rat Insulin ELISA kit, Mercodia Inc., Erling-Holmlund, Sweden) purchased from Shikoku Medical Instruments, Kagawa, Japan.

#### Statistical analysis

Blood glucose (Experiment 1) and plasma glucose and insulin (Experiment 2) were analyzed statistically. For these tests, male and female subject data were combined. The values for analysis were determined before the intake point as well as at 30, 60, 90 and 120 min after intake of each test meal. All measurements are expressed as means  $\pm$  standard error. To examine the significance of differences, Student's paired *t*-test was employed with a level of significance at p < 0.05. Statistical processing was performed using Excel Statistics 2008 (SSRI Co., Ltd., Tokyo, Japan).

#### RESULTS

#### **Experiment 1**

There were no dropouts among the 43 subjects participating in this trial. The glycemic responses for the 3 test meals with sucrose or D-psicose are shown in Table 3. The blood glucose concentrations after each test meal increased over time until 30-60 min and then decreased. The increases in blood glucose concentration after intake of various confections were significantly lower with the simultaneous intake of coffee with D-psicose than that of coffee with sucrose. Statistically significant differences (p<0.05) between sucrose and D-psicose were seen with marshmallow intake after 30 min, fried cookie intake after 30, 60 and 120 min, and chocolate intake after 60 and 90 min. Increments of area under the curve of blood glucose were also lower in each test meal with D-psicose than with sucrose (marshmallow, 5,001 vs. 4,480; fried cookie, 4,002 vs. 2,999; 2,400 vs. 1,736 min·mg/dL, respectively).

#### **Experiment 2**

There were no dropouts among the 9 subjects participation in this trial. The glycemic and insulinemic responses for the 5 test meals containing sucrose or D-psicose are shown in Tables 4 and 5. The plasma glucose and insulin concentrations after each test meal increased over time until 30 min and then decreased. The increases in plasma

glucose concentration were significantly lower after intake of almond jelly and ganache containing D-psicose than those containing sucrose. Statistically significant differences (p<0.05) between sucrose and D-psicose were seen with almond jelly intake after 30, 90 and 120 min and ganache intake after 30, 60 and 90 min. The postprandial plasma glucose concentration did not differ between intake of D-psicose and sucrose containing confections cooked at high temperature (cake, baked and fried cookies). Increments of area under the curve of plasma glucose in the test meals containing D-psicose compared to those containing sucrose were as follows: cake, 3,638 vs. 4,425; almond jelly, 4,437 vs. 3,018; baked cookie, 1,681 vs. 894; fried cookie, 3,638 vs. 4425; ganache, 5,509 vs. 2,823 min · mg/dL, respectively).

The increases in plasma insulin concentration were significantly lower after intake of almond jelly and ganache containing D-psicose than those containing sucrose. Statistically significant differences (p<0.05) between sucrose and D-psicose were seen with almond jelly intake after 30 and 60 min and ganache intake after 30 min. The increases in plasma insulin concentration did not differ among the intake of cake, baked and fried cookies. Increments of area under the curve of plasma insulin in the test meals containing D-psicose compared to those containing sucrose were as follows: cake, 2,313 vs. 2,421; almond jelly, 1,503 vs. 526; baked cookie, 963 vs. 884; fried cookie, 2,421 vs. 2,313; ganache, 2,667 vs. 2,007 min  $\cdot$  mU/L, respectively).

Test meels	Sweeteners	Time after ingestion (min)						
Test means		0	30	60	90	120		
Marshmallow	Sucrose	92.1±2.1	163.1±3.8	147.3±8.6	125.6±7.7	106.1±4.9		
	D -Psicose	90.5±1.5	140.9±4.8*	$146.9 \pm 7.4$	125.2±5.5	$106.2 \pm 1.9$		
Fried cookie	Sucrose	97.2±3.8	143.6±5.6	146.2±4.9	128.2±4.9	111.2±2.7		
	D -Psicose	96.2±1.3	131.8±2.7*	131.3±5.0*	121.5±2.5	104.1±2.6*		
Chocolate	Sucrose	93.6±2.9	114.4±3.5	119.0±3.3	120.9±3.5	106.6±2.3		
	D -Psicose	92.6±1.9	110.6±3.2	108.0±2.4*	110.9±3.3*	$104.9 \pm 2.2$		

**Table 3.** Blood glucose concentrations (mg/dL) after ingestion of test meals with sucrose or D-psicose (Experiment 1).

Values are means  $\pm$  SE for 14-15 subjects. \*p < 0.05, vs. test meals with sucrose (Student's paired *t*-test).

Test meels	Additive sugars	Time after ingestion (min)						
Test means		0	30	60	90	120		
Calza	Sucrose	89.5±3.1	151.9±9.3	121.2±8.3	109.8±5.4	103.2±6.7		
Cake	D -Psicose	83.2±2.7	$146.9 \pm 10.1$	$129.4 \pm 8.2$	112.2±6.0	$100.4 \pm 5.6$		
Almond jelly	Sucrose	72.8±1.7	134.3±6.0	113.2±6.2	$101.6 \pm 5.1$	$107.2 \pm 8.3$		
	D -Psicose	$70.8 \pm 4.9$	117.2±5.8*	$105.0 \pm 9.7$	85.1±2.5*	82.2±6.2*		
Baked cookie	Sucrose	81.5±1.2	103.8±4.6	102.9±4.3	91.2±4.2	86.8±3.3		
	D -Psicose	81.4±2.9	94.8±4.2	$94.4{\pm}4.1$	83.3±4.4	84.4±4.5		
Fried cookie	Sucrose	89.5±3.1	151.9±9.3	121.2±8.3	109.8±5.3	103.2±6.7		
	D -Psicose	83.2±2.7	$146.9 \pm 10.1$	$129.4 \pm 8.2$	112.2±6.0	$100.4 \pm 5.6$		
Ganache	Sucrose	87.7±6.4	$162.6 \pm 5.7$	$147.2 \pm 8.9$	$124.4\pm5.9$	112.8±12.3		
	D -Psicose	86.0±1.2	124.5±4.6*	108.0±6.6*	110.9±4.7*	$104.9 \pm 4.1$		

**Table 4.** Plasma glucose concentrations (mg/dL) after ingestion of test meals with sucrose or D-psicose (Experiment 2).

Values are means  $\pm$  SE for 9 subjects. \*p < 0.05, vs. test meals with sucrose (Student's paired *t*-test).

**Table 5.** Plasma insulin concentrations (mU/L) after ingestion of test meals with sucrose or D-psicose (Experiment 2).

Tast mosls	Additivo sugars	Time after ingestion (min)					
Test means	Additive sugars	0	30	60	90	120	
Calca	Sucrose	4.2±0.7	40.0±7.0	23.3±4.4	21.4±4.2	14.2±4.1	
Cake	D-Psicose	4.0±0.6	32.6±6.4	30.8±6.8	24.0±5.0	14.6±3.8	
Almond jally	Sucrose	9.7±1.7	39.9±6.0	19.6±4.2	16.6±5.1	15.9±8.3	
Annona jeny	D-Psicose	9.6±0.8	19.7±1.5*	13.5±1.0*	12.8±1.2	10.3±2.8	
Dalrad applica	Sucrose	6.7±1.5	$19.8 \pm 4.2$	18.6±3.6	13.1±2.2	8.1±2.4	
Dakeu Cookie	D-Psicose	$6.0{\pm}1.0$	18.6±3.3	17.7±3.3	$10.7 \pm 1.8$	6.9±1.8	
Fried ecolric	Sucrose	4.0±0.6	30.8±6.4	$32.6 \pm 6.8$	$24.0\pm5.0$	14.6±9.8	
FILEU COOKIE	D-Psicose	4.2±0.7	$40.0 \pm 7.1$	23.3±3.8	21.4±4.2	$14.2 \pm 4.1$	
Canaaha	Sucrose	$4.4\pm0.4$	43.9±6.3	$29.9 \pm 5.0$	22.1±7.7	$16.8 \pm 2.9$	
Gallache	D-Psicose	4.6±0.6	31.6±4.6*	23.8±5.0	20.7±4.6	13.8±2.6	

Values are means  $\pm$  SE for 9 subjects. \*p<0.05, vs. test meals with sucrose (Student's paired *t*-test).

# DISCUSSION

The present study suggested that D-psicose is effective for the hypoglycemic response as a functional food material in healthy subjects.

In our previous animal study, suppression of the increase in plasma glucose concentration with D-psicose showed significant decreases when maltose and sucrose were used as substrates, but no significant decreases were observed when glucose and soluble starch were used as substrates [13].

Another animal study proposed that D-psicose inhibited the hydrolysis of maltose by  $\alpha$ -glucosidase prepared from the membrane of the rat small intestine [14]. It follows from these observations that one of the suppressive mechanisms of D-psicose on the elevation of plasma glucose concentration of rats after carbohydrate administration is the inhibition of  $\alpha$ -glucosidase. Suppression of the elevation of plasma glucose concentration in humans with D-psicose was expected when several types of sugars were used as a carbohydrate source. Iida et al. [15] reported dose-dependent effects of D-psicose on suppression of the elevation of plasma glucose and insulin concentration with concurrent administration of maltodextrin and D-psicose in healthy humans. They concluded that D-psicose is efficacious in suppressing of the elevation of blood glucose concentration after eating in humans.

As another hypothetical mechanism for suppression of the increase in plasma glucose concentration, absorbed D-psicose in small intestine, in which D-psicose was estimated to absorb at 25% [18, 19], promoted uptake of glucose in the liver. It has been reported that D-fructose activates glucokinase and reduces plasma glucose concentration after being phosphorylated into fructose 1-phosphate by fructokinase in the liver [20, 21]. A similar mechanism of reducing plasma glucose concentration is also postulated for D-tagatose, an isometric form of D-psicose [22]. The same biochemical pathway as D-fructose and D-tagatose could accordingly enhance glucose tolerance.

The present applied study of D-psicose suggested that hypoglycemic and hypoinsulinemic responses to D-psicose used as raw material in foods were not found in those cooked at high temperature, i.e., cake, baked and fried cookies, in Experiment 2. These confections were made at more than 180°C with relatively long cooking times (dozens of minutes), whereas the others, i.e., almond jelly and ganache, were made at less than 90°C with short cooking times (several minutes). In Experiment 1, D-psicose was hardly heated because it was used as a coffee sweetener. These results suggest that the hypoglycemic effect of D-psicose may be suppressed by cooking at high temperature together with other raw materials.

D-Psicose is a reducing sugar that nonenzymatically glycates the amino groups of proteins or peptides, similar to D-glucose or D-fructose (aminocarbonyl reaction or Maillard reaction) [23-25]. This reaction proceeds through two stages. In the early stage, the sugar reacts with the amino groups of protein/peptide to form a stable Amadori and Heyn's product via a labile Schiff base [26]. In the advance stage, many different complex reactions occur, and consequently, brown, crosslinked fluorescent products are produced [27].

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Amino-carbonyl modification of protein functionality, and the physiological and pathological consequences of protein glycation have been the subject of much investigation. Moreover, amino-carbonyl reaction modifies carbohydrate metabolism [27]. Aminocarbonyl products are hardly digested and absorbed in the mammalian intestine [27]. D-Psicose-protein/peptide conjugates may be excreted into feces without digestion and absorption. As amino-carbonyl reaction is promoted by heating at high temperatures, the disappearance of the hypoglycemic effect of D-psicose may be due to amino-carbonyl reaction with food proteins or peptides. However, detailed studies are required to clarify this mechanism. Care is required when using D-psicose as a food material.

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### REFERENCES

- 1. Cree, G. M. and Perlin, A. S. 1968, Can. J. Biochem., 46, 765.
- 2. Binkley, W. W. 1963, Int. Sugar J., 65, 105.
- Miller, B. S. and Swain, T. 1965, J. Sci. 3. Food Agric., 11, 344.
- Hough, L. and Stacey, B. E. 1966, 4. Phytochemistry, 5, 171.
- Eble, T. E., Hoeksema, H., Boyack, G. A., 5. and Savage, G. M. 1959, Antibio. Chemother., 9, 419.
- Granstrom, T. B., Takata, G., Tokuda, M., 6. and Izumori, K. 2004, J. Biosci. Bioeng., 97, 89.
- Matsuo, T., Suzuki, H., Hashiguchi, M., and 7. Izumori, K. 2002, J. Nutr. Sci. Vitaminol., 48.77.
- 8. Yagi, K. and Matsuo, T. 2009, J. Clin. Biochem. Nutr., 45, 270.
- 9. Ishii, R., Shirai, Y., and Matsuo, T. 2011, J. Clin. Biochem. Nutr., in press.
- 10. Iida, T., Kishimoto, Y., Yoshikawa, Y., Okuma, K., Yagi, K., Matsuo, T., and Izumori, K. 2007, J. Advd. Food Ingred., 10, 10.

- Matsuo, T., Baba, Y., Hashiguchi, M., Takeshita, K., Izumori, K., and Suzuki, H. 2001, J. Clin. Biochem. Nutr., 30, 55.
- 12. Matsuo, T. and Izumori, K. 2006, Biosci. Biotechnol. Biochem., 70, 2081.
- Matsuo, T. 2006, J. Jpn. Soc. Nutr. Food Sci., 59, 191.
- 14. Matsuo, T. and Izumori, K. 2009, J. Clin. Biochem. Nutr., 45, 202.
- Iida, T., Kishimoto, Y., Yoshikawa, Y., Hayashi, N., Okuma, K., Tohi, M., Yagi, K., Matsuo, T., and Izumori, K. 2008, J. Nutr. Sci. Vitaminol., 54, 511.
- Toyoda, Y., Mori, S., Umemura, N., Futamura, Y., Inoue, H., Hata, T., Miwa, I., Murao, K., Nishiyama, A., and Tokuda, M. 2010, Jpn. Pharmacol. Ther., 38, 261.
- 17. Herme, P. 2002, Larousse des desserts, Editions Larousse, Paris.
- Whisler, R. L., Singh, P. P., and Lake, W. C. 1974, Carbohyd. Res., 34, 200.

- 19. Matsuo, T., Tanaka, T., Hashiguchi, M., Izumori, K., and Suzuki, H. 2003, Asia Pac. J. Clin. Nutr., 12, 225.
- Moore, M. C., Cherrington, A. D., Mann, S. L., and Davis, S. N. 2000, J. Clin. Endocrinol. Metab., 85, 4515.
- 21. Shiota, M., Moore, M. C., Galassetti, P., Monohan, M., Neal, D. W., Shulman, G. I., and Cherrington, A. D. 2002, Diabetes, 51, 469.
- 22. Madenokoji, N., Iino, H., Shimizu, T., Hayakawa, J., Sakashita, M. 2003, Jpn. Soc. Clin. Nutr., 51, 21.
- 23. Sun, Y., Hayakawa, S., and Izumori, K. 2004, J. Agric. Food Chem., 52, 1293.
- Sun, Y., Hayakawa, S., Ogawa, M., and Izumori, K. 2005, J. Agric. Food Chem., 53, 10205.
- 25. Sun, Y., Hayakawa, S., and Izumori, K. 2004, J. Food Sci., 69, 427.
- 26. Ames, J. M. 1990, Trends Food. Sci. Technol., 1, 150.
- 27. Ames, J. M. 1998, Food Chem., 62, 431.